

Version with Markings to Show Changes Made

The amended paragraphs indicate deletions by ~~strikeout~~ and insertions by underlining.

In the specification, page 3, second full paragraph.

*Ballabio et al.* (1991), disclose a ~~single-tube multiplex~~  
~~allele-specific~~ single-tube, multiplex allele-specific PCR test using  
two different dye-tagged fluorescent primers for detection of the  
 $\Delta F508$  cystic fibrosis mutation.

In the specification, page 11, first full paragraph.

The primers must also be designed so that the size of the  
resulting amplification products differ in length, thereby facilitating  
assignment of alleles to individual loci during detection.  
Inappropriate selection of primers can produce several undesirable  
effects such as lack of amplification, amplification at multiple sites,  
primer dimer formation, undesirable interaction of primer  
sequences from different loci, production of alleles from one locus  
which overlap with alleles from another, or ~~requirement~~ the need  
for amplification conditions or protocols for the different loci  
which are incompatible in a multiplex. The synthesis of the  
primers is conducted by procedures known to those skilled in the  
art.

In the specification, page 18, third full paragraph.

In this example, a DNA template was amplified at the  
individual loci HUMCSF1PO, HUMTPOX, HUMTH01, and  
HUMVWFA31 simultaneously in a single reaction vessel. The  
PCR amplifications were performed in 25 $\mu$ l volumes using 25ng  
template, 0.04U *Taq* DNA Polymerase/ $\mu$ l, 1x STR Buffer (50mM  
KCl, 10mM Tris-HCl (pH 9.0 at 25°C), 0.1% Triton X-100, 1.5mM  
MgCl<sub>2</sub> and 200 $\mu$ M each of dATP, dCTP, dGTP and dTTP), and  
using a Thermal Cycler 480 (Perkin Elmer Cetus). Amplification  
protocol 1, as described in Example 1, was employed. Eight

amplification primers were used in combination, including 1 $\mu$ M each HUMCSF1PO primer 2 [SEQ. ID. 5] [SEQ. ID. 6] and fluorescein-labeled primer 1 [SEQ. ID. 5], 0.15 $\mu$ M each HUMTPOX primer 1 [SEQ. ID. 29] and fluorescein-labeled primer 2 [SEQ. ID. 30], 0.2 $\mu$ M each HUMTH01 primer 2 [SEQ. ID. 28] and fluorescein-labeled primer 1 [SEQ. ID. 27], and 1 $\mu$ M each HUMVWFA31 primer 1 [SEQ. ID. 31] and fluorescein-labeled primer 2 [SEQ. ID. 32].